

## Intravital imaging of secondary axon death after spinal cord injury

### Neuroscience

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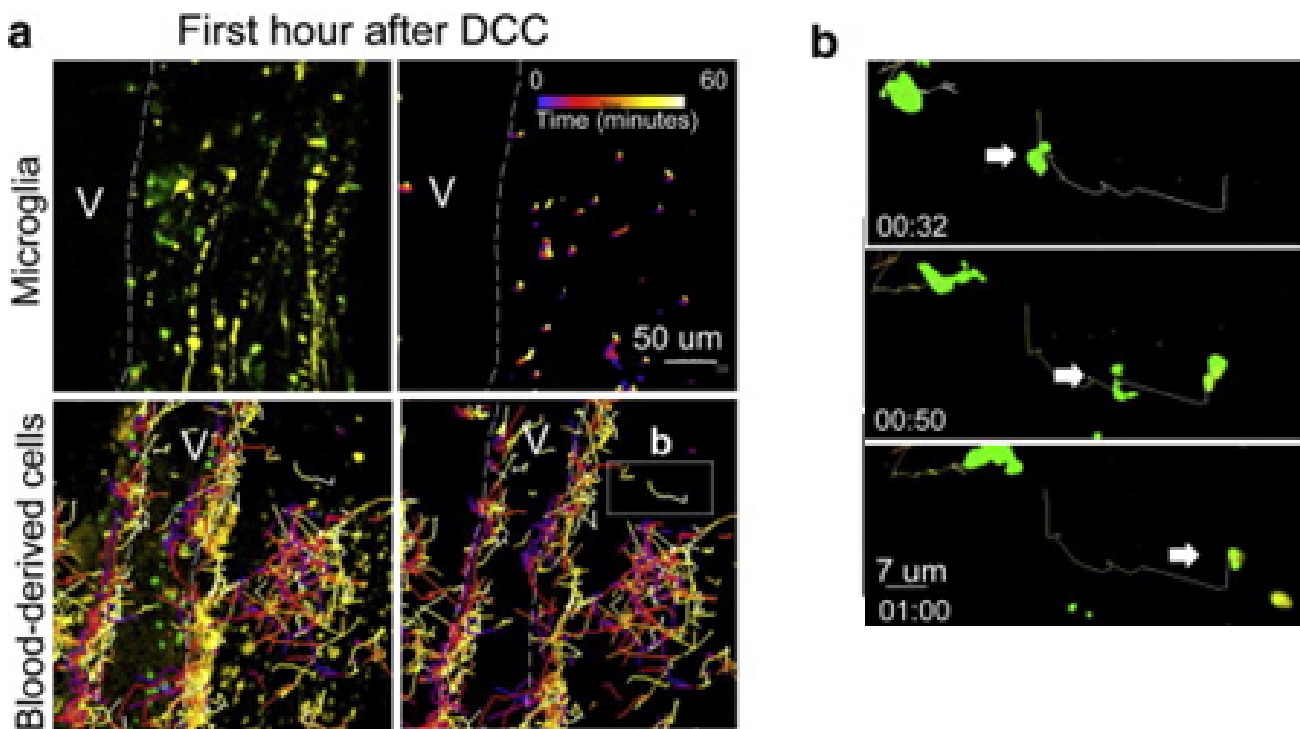
Researchers led by Drs. Jerry Silver and Alex Y. Huang at Case Western Reserve University School of Medicine used Imaris software together with time-lapse multiphoton microscopy to visualize the axonal injury that occurs after spinal cord trauma in living mice.

When the spinal cord is injured, an initial dieback of axons during the first few hours is followed by a secondary phase of axon death over the following weeks. The researchers wanted to learn more about this secondary phase because understanding the mechanisms that cause axonal injury to increase during this phase could lead to treatments that decrease additional functional deficits that occur as the lesion size and damage increases.

“Currently there are few, if any, pharmacological treatments that help patients maintain any residual function immediately after a spinal cord an injury,” says Dr. Teresa Evans, a member of the research team. “Decreasing this secondary injury will likely play a huge role in the treatment of patients using combinatorial approaches to treatment, along with immediate surgical interventions and regenerative therapies that are also currently under investigation.”

### Real-time imaging in 3-D

During the secondary period of dieback, axons approaching the lesion lose parts of their membrane as the size of the lesion increases and it is infiltrated by blood-derived immune cells, including macrophages. To analyze this immune process, the researchers acquired intravital real-time multiphoton microscopy images of the interactions of GFP-labeled macrophages and GFP-labeled microglia with YFP-labeled axons in a mouse model of spinal cord injury. On the day of spinal injury and 2, 4, 5 and 8 days after, the researchers acquired 3-D images covering the top ~150 microns of the spinal column with a z-step size of 3 microns.

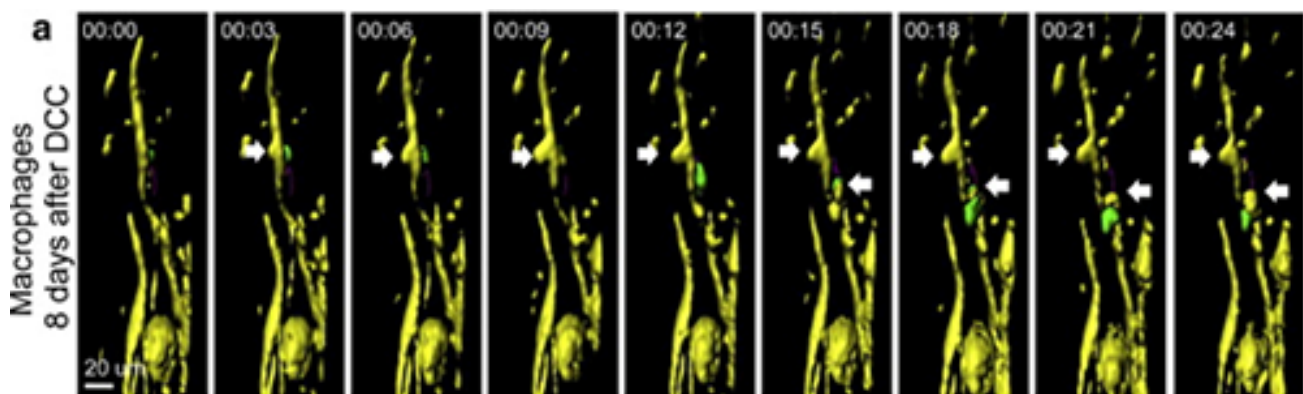


Blood-derived CX3CR1+ macrophages have greater average speed and displacement than resident microglia. a) Immediately after dorsal column crush (DCC) lesion, Cx3cr1+/GFP microglia move only very short distances, while blood-derived cells traversed over great distances within the center of the lesion. Cell tracks are shown as color-coded time stamps (blue: time = 0 min; yellow: time = 60min; gradient shown in the figure). Both tracks and fluorescence are shown in images on the left, and only tracks in images on the right. Scale bar= 50 microns. Total time for tracking is 1 h.

b) High-magnification sequential images of one macrophage's track and movement in a. Scale bar=7 microns. Time stamp: h:min. Total time shown is 28 min. Reprinted from Evans et al. *Exp Neurol.* 254:109-20.

"For us, the ability to work with very large, three dimensional, multichannel data sets was a challenge, and Imaris allowed us to do this," says Dr. Evans "Intravital imaging in live animals results in huge datasets, and being able to work with these in a reliable manner was integral to the success of these studies."

The researchers used the Imaris Spots Function for automated and manual counting and tracking of multiple well-defined cells in three-dimensional space. They also used Imaris to conveniently examine the time series of z-stacks to observe changes in 3-D structures over time. With Imaris, they could cut these stacks at different angles and to different thicknesses to easily view movement of structures within the stacks over time.



Axonal destruction occurs in response to contact only with Cx3cr1+/GFP blood-derived cells and not Cx3cr1+/GFP microglia. a) Example of contact between a Thy-1CFP/+ axon (yellow) and a Cx3cr1+/GFP blood-derived cell (green) resulting in axonal "blebbing" (white arrows) on day 5 after DCC injury. Blebbing occurred in two locations of the axon (yellow). Reprinted from Evans et al. *Exp Neurol.* 254:109-20.

Overall, the analysis revealed that blood-derived macrophages but not microglia facilitate secondary axonal dieback that occurs in traumatic spinal cord injury. The researchers continue to use Imaris for many ongoing projects, including 3-D tracking of cell movement in many different tissues such as the brain, bone marrow, lymph node and other areas.

This movie shows an example of axonal thinning in an animal imaged 5 days after initial injury. Over a period of 2 h, the yellow axon thins to the point of breaking at two places along the axon, both in contact with a green Cx3cr1+/GFP microglia or macrophage. Both break points are at points of contact with a Cx3cr1+/GFP cell. Playback speed = 300 x. Second, we show that even at 22 days after injury, small portions of membrane can be lost from the ends of axons when in contact with a Cx3cr1+/GFP cell. Here, axonal cleavage is shown near the end of the axon, marked by a blue arrow. From Evans et al. *Exp Neurol.* 254:109-20.

**Research Paper:** [Evans TA, Barkauskas DS, Myers JT, Hare EG, You JQ, Ransohoff RM, Huang AY, Silver J. 2014. High-resolution intravital imaging reveals that blood-derived macrophages but not resident microglia facilitate secondary axonal dieback in traumatic spinal cord injury. \*Exp Neurol.\* 254:109-20.](#)